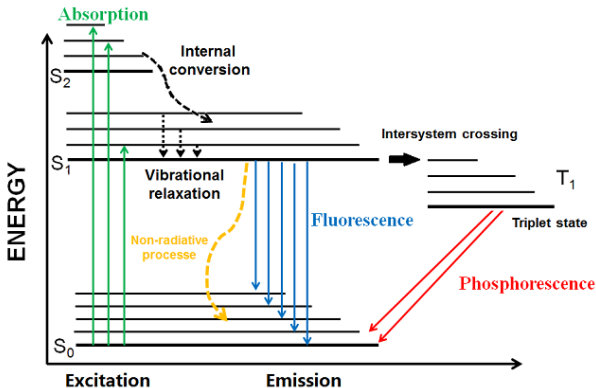


# Ringvorlesung Biophysik 3

## Theoretische Aspekte der Photobiologie

AK Burghardt (Institut für Physikalische & Theoretische Chemie)





# Theoretical Chemistry of Complex Systems

AK Burghardt



chemical + biological + material processes

chemistry  
“bottom-up”:

↑  
electrons + nuclei



# Theoretical Chemistry of Complex Systems

AK Burghardt



quantum density

$$\rho \longleftrightarrow \psi^* \psi$$

wave function

classical probability  
distribution

$$\begin{array}{c} \updownarrow \\ P \end{array}$$



# Theoretical Chemistry of Complex Systems

AK Burghardt



## Teaching:

Theoretische Chemie I (Bachelor) – Burghardt

Theoretische Chemie II + Praktikum (Bachelor/Master) – Burghardt

Theoretical Photochemistry (Master) – Burghardt

Mathematische Methoden I (Bachelor) – Hegger

Mathematische Methoden II (Bachelor) – Hegger

Mathematische Methoden III (Bachelor, Master) – Hegger

+ MD-QC Praktikum (Bachelor<sup>1</sup>) – Burghardt, Hegger

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<sup>1</sup>Wahlpflichtmodul “Computational Chemistry”

# Topics

## 1 Photochemistry

Quantum Effects in Biological Systems

Photochemistry: Quantum Mechanics at Work

Quantum (QM) or Quantum/Classical (QM/MM-MD)?

# Topics

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Quantum Effects in Biological Systems

Photochemistry: Quantum Mechanics at Work

Quantum (QM) or Quantum/Classical (QM/MM-MD)?

## 2 Modeling of Photoinduced Uncaging @CLiC

Can Theory Predict the Best Cages?

Spectroscopy: What's the Best Way to Prepare your System?

How to Tackle Multiple Time Scales?

# Topics

- 1 Photochemistry
  - Quantum Effects in Biological Systems
  - Photochemistry: Quantum Mechanics at Work
  - Quantum (QM) or Quantum/Classical (QM/MM-MD)?
- 2 Modeling of Photoinduced Uncaging @CLiC
  - Can Theory Predict the Best Cages?
  - Spectroscopy: What's the Best Way to Prepare your System?
  - How to Tackle Multiple Time Scales?
- 3 Photoregulation of DNA & RNA
  - Conformational Control using Covalently Attached Photoswitches
  - QM/MM Simulations
  - MM-MD Simulations

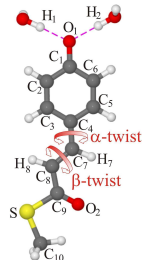
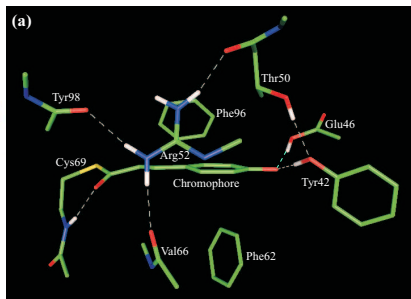
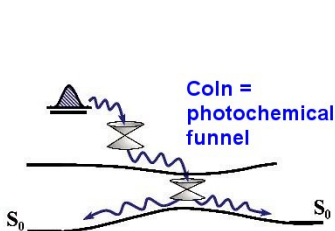
## Why Worry about Quantum Effects?

- Most properties/processes in biological & material systems can be taken to belong to the classical limit ( $\lambda_{dB} \ll L$ , classical Wigner limit, ...)
- Hence, MD (= Molecular Dynamics) simulations are extensively used: classical-mechanical evolution of all nuclear degrees of freedom on the lowest (electronic ground state) Born-Oppenheimer (BO) surface
- However, some quantum effects are important:
  - tunneling (proton transfer, electron transfer)
  - light-induced processes: coherent superpositions
  - nonadiabatic (“non-BO”) dynamics
- Methods are needed for quantum dynamics in many dimensions
  - approximate wavefunction and density operator methods
  - semiclassical approaches
  - mixed quantum-classical hybrid approaches



# Photoactive Proteins: Bio-Photochemistry

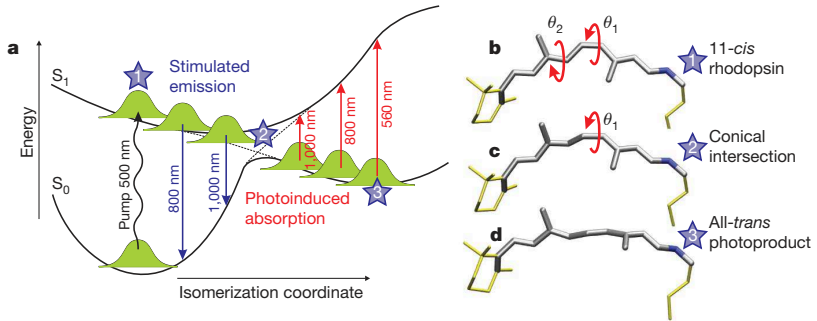
Example PYP = Photoactive Yellow Protein



Gromov, Burghardt, Köppel, Cederbaum, J. Phys. Chem. A **115**, 9237 (2011), JACS **129**, 6798 (2007)

- photochemistry determined by conical intersection (CoIn) topology
- local environment significantly influences  $\alpha$  vs.  $\beta$  twist
- interplay of steric effects / electrostatics / H-bonds / fluctuations
- excited-state lifetime:  $\sim 700$  fs (in solution:  $\sim 10$  ps)

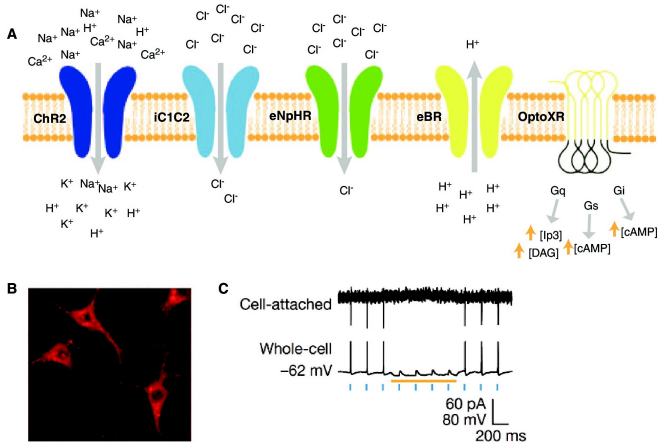
# Bio-Photochemistry: Retinal/Rhodopsin



Polli et al., Nature 467, 440 (2010)

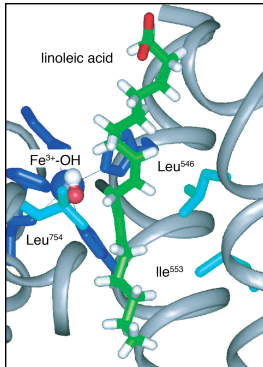
- isomerisation within  $\sim 200$  femtoseconds (but 5 picoseconds in solution)
- isomerisation happens in a volume conserving fashion

# From Bio-Photochemistry to Optogenetics



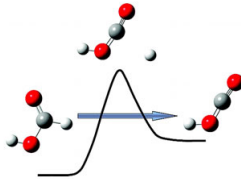
Guru et al., *Int. J. Neuropsychopharmacology* 1-8 (2015)

# Quantum Tunneling in Enzymes



Meyer, PNAS 105, 1146 (2008)

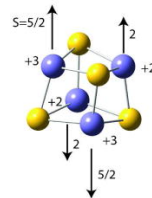
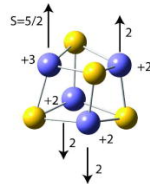
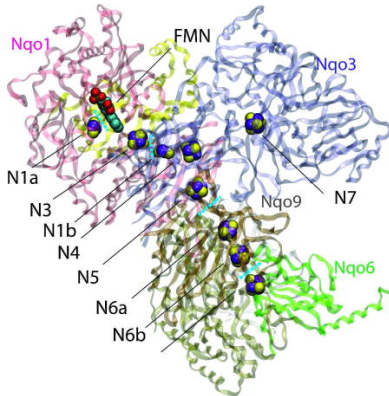
"Enzyme structure and dynamics affect hydrogen tunneling:  
The impact of a remote side chain (I553) in soybean lipoxygenase-1"



- "Did enzymes evolve to capitalize on quantum tunneling?" The Scientist, 2005
- "Our present findings on hydrogen transfer under physiological conditions cannot be explained without invoking both quantum mechanics and enzyme dynamics" Klinman, Nature 1999
- "Taking Ockham's razor to enzyme dynamics and catalysis": no need to go beyond Transition State Theory (TST) + tunneling corrections

Glowacki et al., Nature Chem. 2012

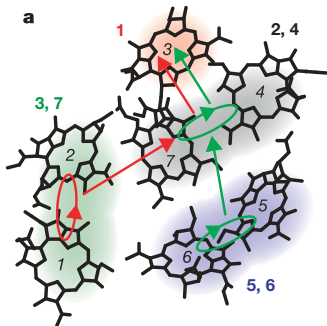
# Electron Tunneling: Respiratory Chains



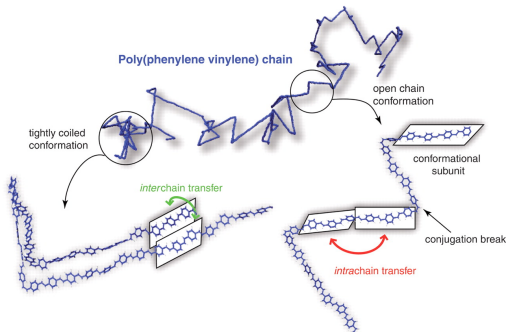
“Quantum Electron Tunneling in Respiratory Complex I1”, Hayashi, Stuchebrukhov, J. Phys. Chem. B 115, 5354 (2011)

- “electronically wired” iron-sulfide (FeS) clusters
- quantum interference resulting from multiple tunneling pathways

# Coherent Photoinduced Energy and Charge Transport in Biological and Material Light-Harvesting Systems



Lee, Cheng, Fleming, Science 316, 1462 (2007)

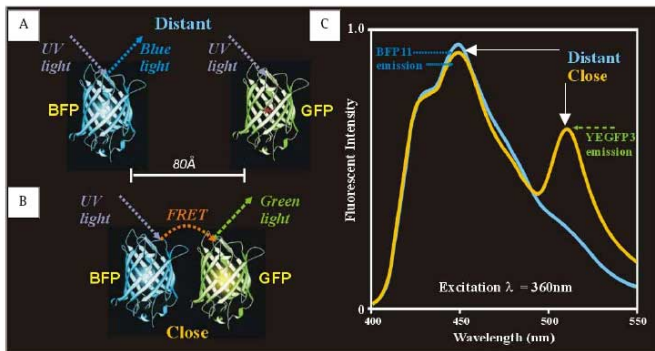


Collini, Scholes, Science 323, 369 (2009)

- one would expect an extremely rapid dephasing (decoherence):  $\tau_{\text{dec}} < 50$  fs
- but observed coherence lifetimes are  $\sim 300$  fs to 1-2 ps (or more)

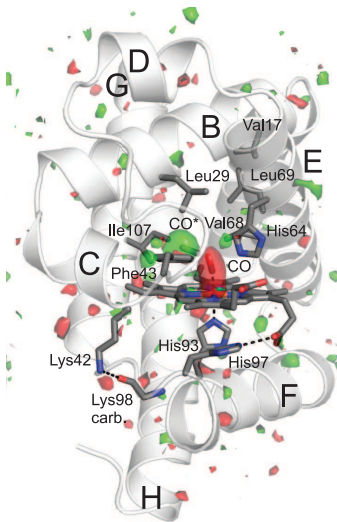
# Fluorescence Resonance Energy Transfer (FRET)

E.g., Fluorescence Resonance Energy Transfer (FRET) between fluorescent proteins: calculate FRET rates  $\propto r_{DA}^{-6}$



**Figure 1.** The use of GFP variants to produce FRET. In A, Distant; UV light excites BFP (donor fluorophore) to emit blue light (peak emission = 450nm), but GFP (acceptor) is not close enough to draw energy from the excited donor. In B, Close; proximity of GFP to BFP allows non-radiative energy transfer, the stimulated BFP exciting GFP to fluoresce green (peak emission = 510nm). Figure 1C shows the expected emission spectra when the GFP fluorophores are either close or distant.

# Photoinduced CO Ligand Dissociation from Myoglobin



## Direct observation of ultrafast collective motions in CO myoglobin upon ligand dissociation

Thomas R. M. Barends,<sup>1\*</sup> Lutz Foucar,<sup>1</sup> Albert Ardevol,<sup>2</sup> Karol Nass,<sup>1</sup> Andrew Aquila,<sup>2</sup> Sabine Botha,<sup>1</sup> R. Bruce Doak,<sup>1</sup> Konstantin Falahati,<sup>4</sup> Elisabeth Hartmann,<sup>1</sup> Mario Hilpert,<sup>2</sup> Marcel Heinz,<sup>2,4</sup> Matthias C. Hoffmann,<sup>2</sup> Jürgen Köfinger,<sup>2</sup> Jason E. Koglin,<sup>2</sup> Gabriela Kovacsova,<sup>1</sup> Mengning Liang,<sup>2</sup> Despina Milathianaki,<sup>2</sup> Henrik T. Lemke,<sup>2</sup> Jochen Reinstein,<sup>1</sup> Christopher M. Roome,<sup>1</sup> Robert L. Shoeman,<sup>1</sup> Garth J. Williams,<sup>2</sup> Irene Burghardt,<sup>4</sup> Gerhard Hummer,<sup>2</sup> Sébastien Boutet,<sup>2</sup> Ilme Schlichting<sup>4\*</sup>

Science, 350, 445 (2015)



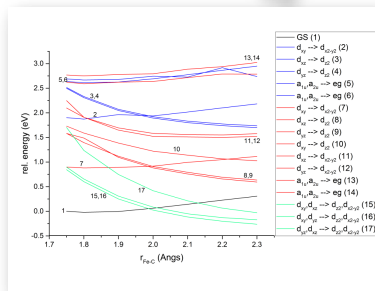
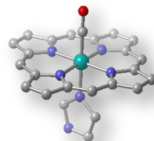
## PHOTODISSOCIATION OF MYOGLOBIN

carboxy-hemoproteins are among the most studied systems to evaluate (collective) protein dynamics that determine structure and reactivity

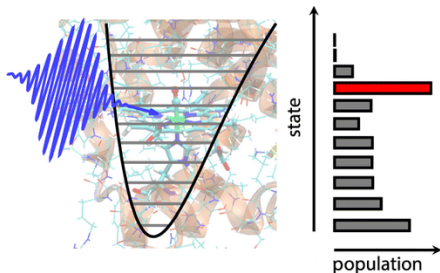
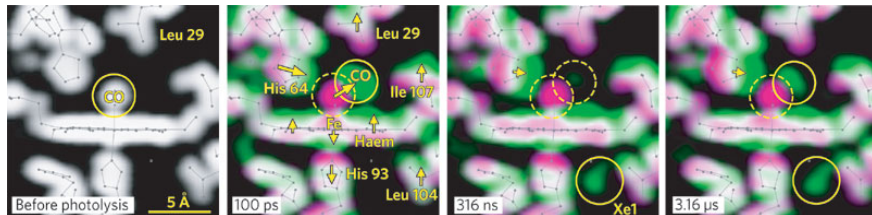
quantum mechanical *ab initio* calculations at TD-DFT and CASSCF/CASPT2 level reveal complex setting of excited states

large manifolds of singlet, triplet and quintet states within range of experimental excitation

interaction based on vibronic coupling and/or spin-orbit coupling may cause overall ultrafast dissociation within < 100 fs

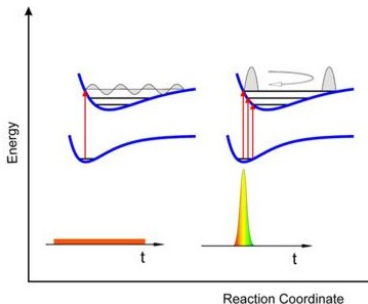


## Quantum Oscillators in Highly Specific Environments



- CO-myoglobin complex
- photoinduced CO dissociation from myoglobin
- state-specific excitation feasible
- relaxation/decoherence depend critically on local environment

# Light Pulses Create Excited-State Wavepackets



left: CW excitation,  
right: pulsed excitation

<http://www.uni-heidelberg.de/fakultaeten/chemgeo/pci/motzkus/research/wavepackets.html>

- wavepacket = superposition of eigenstates

$$\psi(x, t) = \sum_n c_n \phi_n(x) e^{-iE_n t / \hbar}$$

non-stationary state (in contrast to a single eigenstate)

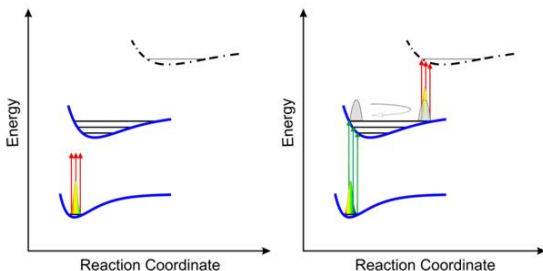
- solve the time-dependent Schrödinger Equation:

$$i\hbar \frac{\partial \psi}{\partial t} = \left( \hat{H}_{\text{mol}} - E(r, t) \hat{\mu} \right) \psi$$

where  $\hat{H}_{\text{mol}}$  is the molecular Hamiltonian and  $\hat{\mu}$  is the dipole operator

# Femtochemistry: Watching Wavepackets in Real Time

Nobel Prize Ahmed Zewail 1999



- nonlinear optical spectroscopy
- wavepacket dynamics in electronically excited states: quantum coherence
- theoretical interpretation: explicit simulation needed!

<http://www.uni-heidelberg.de/fakultaeten/chemgeo/pci/motzkus/research/wavepackets.html>

Pump-Probe Spectroscopy

Theory: **electronic structure** (→ potential surfaces) + **quantum dynamics**

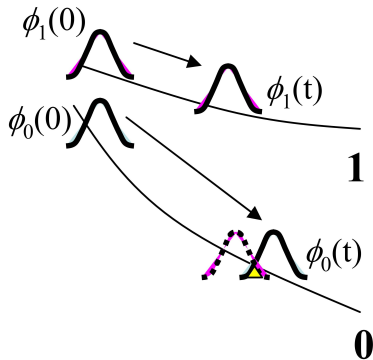
# Quantum Coherence – and Decoherence

$$|\psi(t)\rangle = c_0(t)|0\rangle|\phi_0(t)\rangle + c_1(t)|1\rangle|\phi_1(t)\rangle$$

electronic coherence:

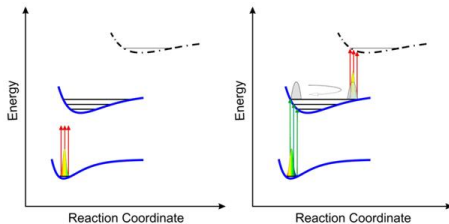
$$\begin{aligned}\rho_{01}(t) &= \text{Tr}[|0\rangle\langle 1|\hat{\rho}(t)] \\ &= \langle 1|\hat{\rho}(t)|0\rangle = c_1^*(t)c_0(t)\langle\phi_1(t)|\phi_0(t)\rangle\end{aligned}$$

- coherence  $\propto$  overlap of nuclear wavefunctions
- typical decoherence times:  $\sim 30$  fs
- loss of coherence cannot be captured by a classical trajectory picture



picture: P. Rossky et al.

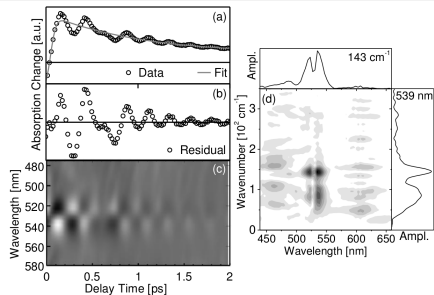
# Vibrational Coherence & Coherence Transfer



## Pump-Probe spectroscopy

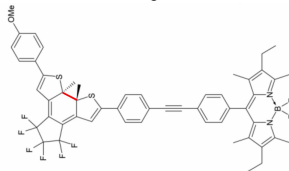
<http://www.uni-heidelberg.de/fakultaeten/chemgeo/pci/motzkus/research/wavepackets.html>

- vibrational coherence  
↔ wavepacket motion
- coherence transfer due to coupled vibrations

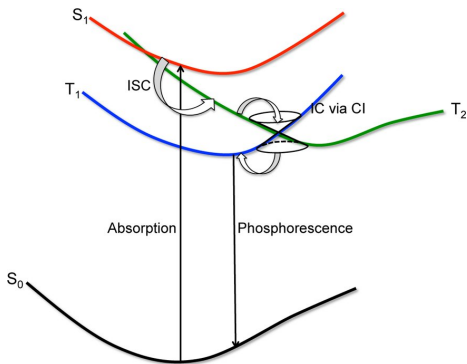


## Vibrational coherence transfer in an electronically decoupled molecular dyad

Schweighöfer et al., Scientific Reports (2015)

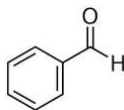


## Even in Small Systems, Excited-State Dynamics is Complex



Ou, Subotnik, J. Phys. Chem. C. 117, 19839 (2013)

- Example: benzaldehyde



- ISC = intersystem crossing
- IC = internal conversion
- CI = conical intersection
- ISC time scale:  
 $\sim 10^{-9}$  s – “untypically” fast!
- phosphorescence lifetime  
 $\sim 2 \times 10^{-3}$  s

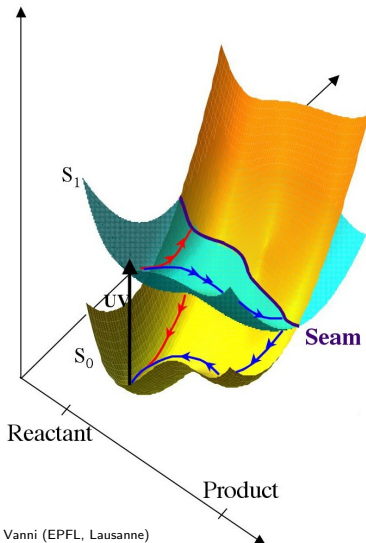
- state-to-state couplings needed
- characterize special topologies: conical intersections
- identify most relevant coordinates

# Beyond Born-Oppenheimer



- “non-adiabatic” dynamics
- quantum dynamical description required
- mean-field (Ehrenfest) and surface-hopping approaches often insufficient

Potential  
Energy



graphics: Stefano Vanni (EPFL, Lausanne)



## What Do We Need To Calculate?

electronic structure :

$$H_{\text{el}}\psi_n^{\text{el}}(r, R) = E_n(R)\psi_n^{\text{el}}(r, R)$$

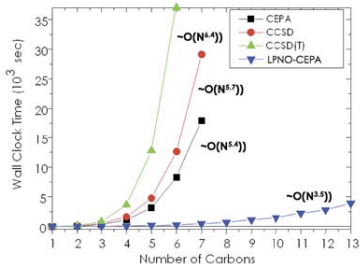
- high-level methods (CC2, ADC(2), CASSCF/CASPT2, MRCI)
- density functional methods (DFT, TDDFT)
- quantum mechanics / molecular mechanics (QM/MM) hybrid methods
- linear scaling methods

quantum dynamics :

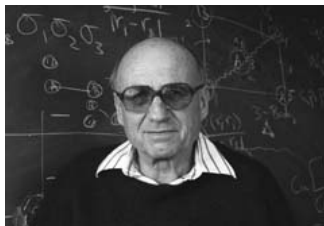
$$i\hbar\psi_n^{\text{nuc}}(R) = (T_{\text{nuc}} + E_n(R))\psi_n^{\text{nuc}}(R)$$

- multiconfigurational methods (MCTDH)
  - mixed quantum-classical methods
  - reduced-dimensional descriptions (effective modes, mesoscopic descriptions)
  - Markovian and non-Markovian master equations
- method development!

## Exponential Scaling Problem



Standard methods: exponential scaling  
 → limit reached already for 6 carbon atoms



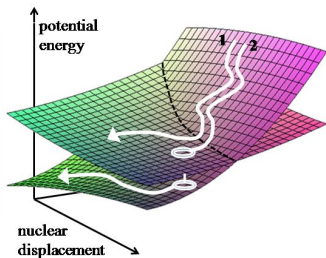
“I cannot foresee any technological progress which should make such calculations feasible.”

*Walter Kohn, Nobel Lecture 1998*

- here, electronic structure example – but same problem for quantum dynamics!
- development of approximate methods is of key importance

## Various Approximate Approaches

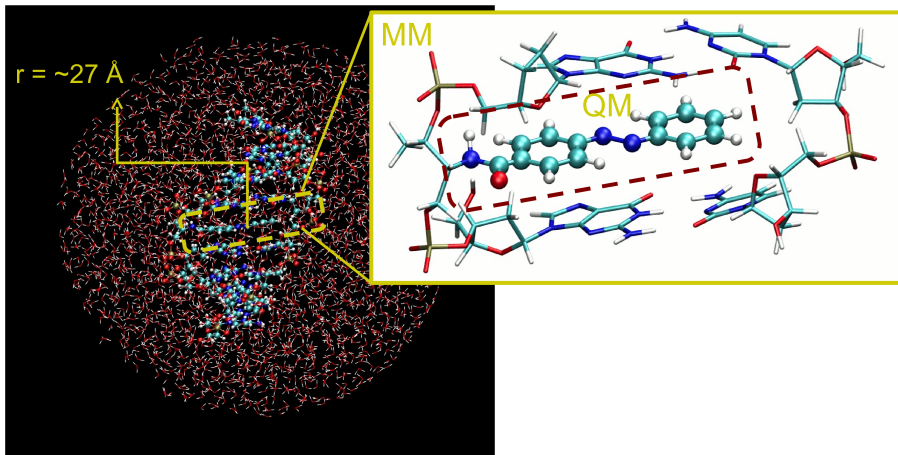
- ① *On-the-fly* Quantum-Mechanics/Molecular Mechanics (QM/MM) + Surface Hopping (SH) – Example: Photocontrol of DNA and RNA
- ② Parametrized Hamiltonians + Quantum Dynamics (QD) – Example: Elementary Events in Organic Photovoltaics



Surface Hopping (SH):  
stochastic trajectories mimic wavepacket  
motion on coupled potential surfaces

## QM/MM simulations

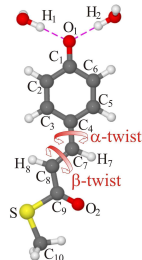
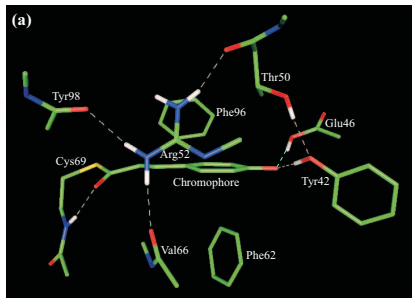
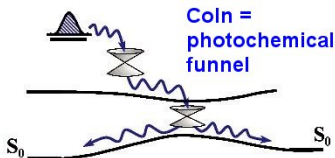
2013 Nobel Prize for Martin Karplus/Michael Levitt/Arieh Warshel



various approaches: NEWTON-X, MNDO/Chemshell, MOPAC (Pisa version), ...

# Photoactive Proteins: Bio-Photochemistry

Example PYP = Photoactive Yellow Protein

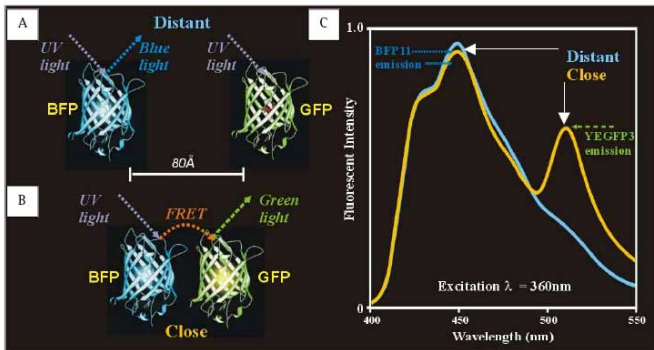


Gromov, Burghardt, Köppel, Cederbaum, J. Phys. Chem. A **115**, 9237 (2011), JACS **129**, 6798 (2007)

- photochemistry determined by conical intersection (CoIn) topology
- local environment significantly influences  $\alpha$  vs.  $\beta$  twist
- interplay of steric effects / electrostatics / H-bonds / fluctuations
- excited-state lifetime:  $\sim 700$  fs (in solution:  $\sim 10$  ps)

## Sometimes Rate Theories are Sufficient . . .

E.g., Fluorescence Resonance Energy Transfer (FRET) between fluorescent proteins: calculate FRET rates  $\propto r_{DA}^{-6}$



**Figure 1.** The use of GFP variants to produce FRET. In A, Distant; UV light excites BFP (donor fluorophore) to emit blue light (peak emission = 450nm), but GFP (acceptor) is not close enough to draw energy from the excited donor. In B, Close; proximity of GFP to BFP allows non-radiative energy transfer, the stimulated BFP exciting GFP to fluoresce green (peak emission = 510nm). Figure 1C shows the expected emission spectra when the GFP fluorophores are either close or distant.

# Topics

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Quantum Effects in Biological Systems

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## 2 Modeling of Photoinduced Uncaging @CLiC

Can Theory Predict the Best Cages?

Spectroscopy: What's the Best Way to Prepare your System?

How to Tackle Multiple Time Scales?

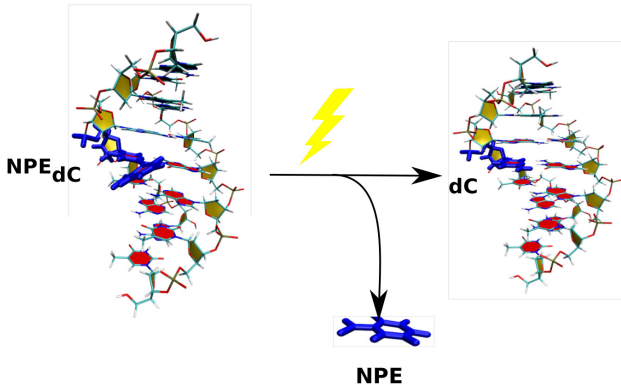
## 3 Photoregulation of DNA & RNA

Conformational Control using Covalently Attached Photoswitches

QM/MM Simulations

MM-MD Simulations

## Photocontrol by Uncaging

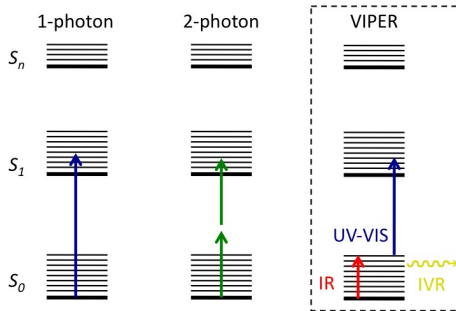


- Graduate School **CLiC** = Complex scenarios of Light Control





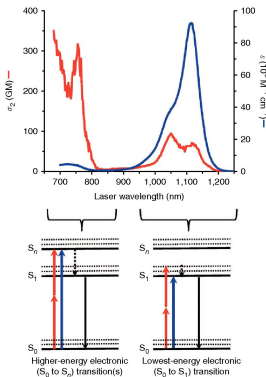
# Spectroscopy: What's the Best Way to Prepare Your System?



Criteria:

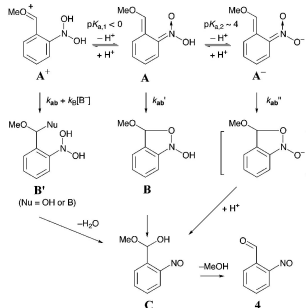
- **absorption efficiency** (e.g., single-photon vs. two-photon)
- **spatial resolution:** two-photon induced fluorescence decrease as the fourth power of the distance from the focal plane (“emission from a single point”)
- **frequency-domain selectivity:** e.g., higher selectivity in the infrared domain as used in the VIPER experiment
- design of **sequential (two-color)** uncaging systems

# Calculation/Prediction of Uncaging Efficiencies (e.g., *o*-Nitrobenzyl Caging Groups)



Drobizhev et al., Nat. Meth. 8, 393 (2011)

**Scheme 4:** Revised Mechanism for the Thermal Reactions of the Primary Photochemical *aci*-Transients A Formed from 1 in Aqueous Solution.<sup>2</sup>

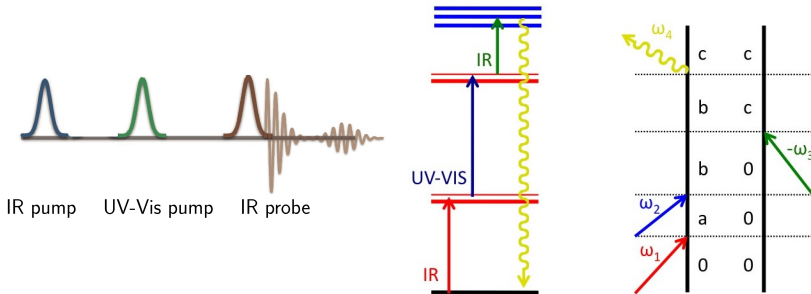


Il'ichev et al., J. Am. Chem. Soc. 126, 4581 (2004)

- efficiency =  $\epsilon \cdot \Phi$  (1-photon) or  $\delta \cdot \Phi$  (2-photon)
- intrinsic quantum yields ( $\Phi$ ): reaction paths, intermediates required

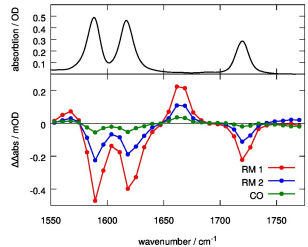
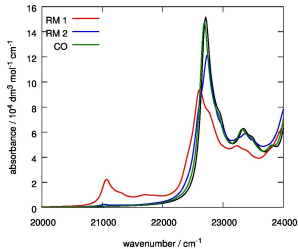
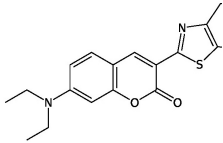
## Spectroscopies can be Sophisticated ...

### A4: Vibrationally Promoted Electronic Resonance (VIPER) 2D spectroscopy



- mixed IR/UV-Vis pulse sequence
- shaped femtosecond IR pulses
- calculate optical response functions

## Simulation of VIPER experiment

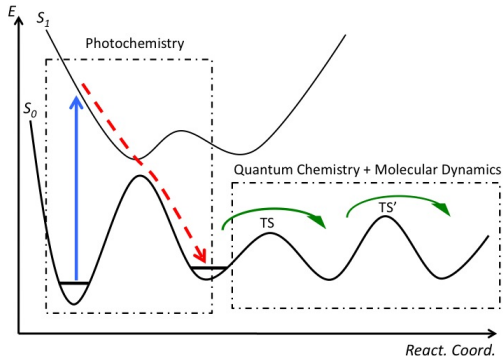


- first step: calculate absorption spectrum with vibrational pre-excitation

$$\alpha(\omega) = \frac{4\pi^2\omega_I}{3\hbar c} \sum_n |\langle \psi_n^E | \hat{\mu} | \psi_0^G \rangle|^2 \delta(\omega_I - \omega_n) = \frac{2\pi\omega_I}{3\hbar c} \int_{-\infty}^{\infty} dt \langle \phi_E(0) | \phi_E(t) \rangle e^{i\omega t}$$

- complementary time-independent and time-dependent approaches (full quantum approach needed, for hundreds of normal modes!)
- identify the modes that are best suited for VIPER effect

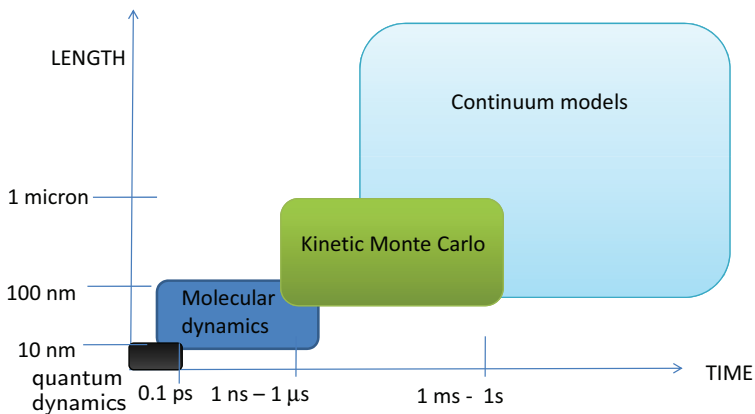
## How to Tackle Multiple Time Scales?



Hierarchy of methods:

- **quantum dynamics**: femtoseconds to picoseconds
- classical **molecular dynamics (MD)**: picoseconds to nano-/microseconds
- **kinetic models**: microseconds to seconds (and longer)

## Hierarchy of Time and Length Scales



- Coarse-graining techniques can bridge length scales

# Topics

## 1 Photochemistry

Quantum Effects in Biological Systems

Photochemistry: Quantum Mechanics at Work

Quantum (QM) or Quantum/Classical (QM/MM-MD)?

## 2 Modeling of Photoinduced Uncaging @CLiC

Can Theory Predict the Best Cages?

Spectroscopy: What's the Best Way to Prepare your System?

How to Tackle Multiple Time Scales?

## 3 Photoregulation of DNA & RNA

Conformational Control using Covalently Attached Photoswitches

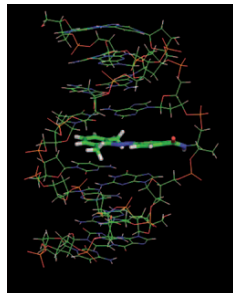
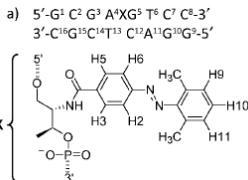
QM/MM Simulations

MM-MD Simulations

## Azobenzene + DNA

- experiments by Asanuma & co: light-triggered conformational changes
- cis/trans melting temperatures are a sensitive function of substituents:
  - 2',6'-dimethyl-*trans*-azobenzene slightly stabilizing
  - 2',6'-dimethyl-*cis*-azobenzene destabilizing
  - p-<sup>i</sup>Pr-*trans*-azobenzene destabilizing
  - p-<sup>i</sup>Pr-*cis*-azobenzene slightly destabilizing

Duplex	Temp. [C]		ΔT
CXG/GC	Trans	Cis	
Native	47,7	47,7	
Azo	48,9	43,2	5,7
2',6'-Me-Azo	50,9	36,3	14,6

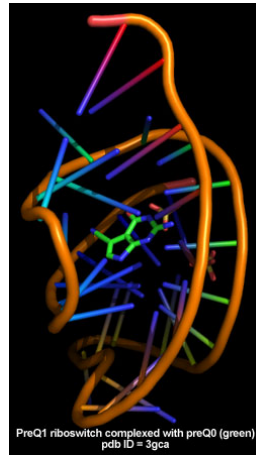


Asanuma and collaborators, Chem. Eur. J. 2009, Nucleic Acids Symposium Series 2007



## RNA: e.g., Riboswitch-Ligand Complexes

- controlled riboswitch (de-)activation by small ligand species (e.g., metal ions, amino acids)
- recent works by Asanuma/Heckel: **photoinduced** activation by photoswitches (azobenzene, spiropyrane)
- characterization of RNA folding intermediates
- **SFB 902** “Molecular mechanisms of RNA based regulation”



Spitale et al., J. Biol. Chem. 284, 11012 (2009)

# How Does a Photoswitch Control DNA / RNA Unfolding?

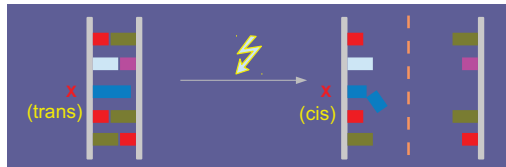
Combine four approaches:

- 1 characterization of chromophore: QM (electronic structure)
- 2 ultrafast photoswitch dynamics (fs-ps scale): QM/MM + MD simulations including RNA
- 3 ps-ns time scale: MD
- 4  $\mu$ s-s time scale: kinetic modeling: e.g., Markov State Models

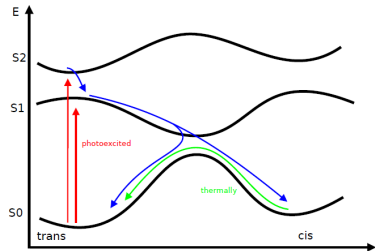
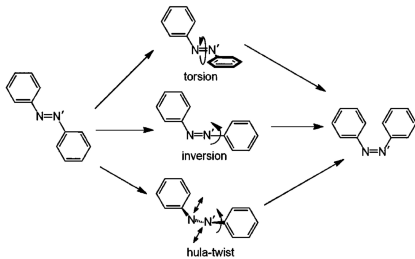
QM = Quantum Mechanics

MM = Molecular Mechanics

MD = Molecular Dynamics



# Azobenzene Switch



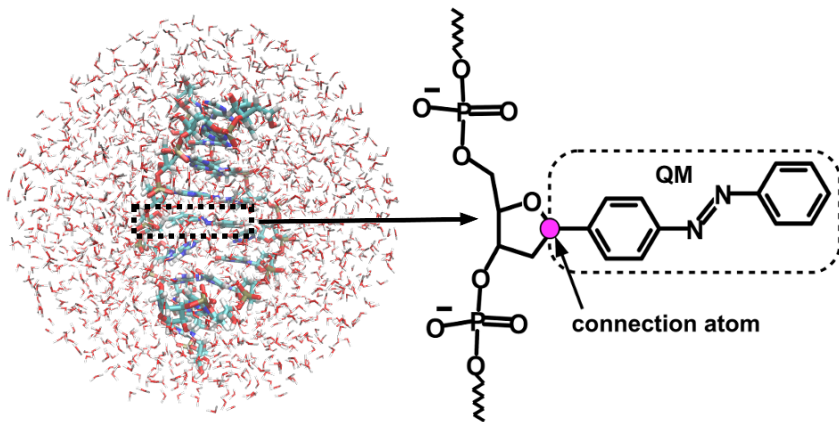
Quick et al., J. Phys. Chem. B, 2014, 118, 8756

- more precise picture: **conical intersection** (“photochemical funnel”)
- **ultrafast** (fs-ps scale) decay of electronically excited states
- several **competing pathways**: isomerization, rotation, “hula-twist”
- picture can be modified by **substituent effects, environmental constraints**

“Femtosecond spectroscopy reveals huge differences in the photoisomerization dynamics between azobenzenes linked to polymers and azobenzenes in solution”, Bahrenburg et al., PCCP 16, 11549 (2014)

## QM/MM simulations

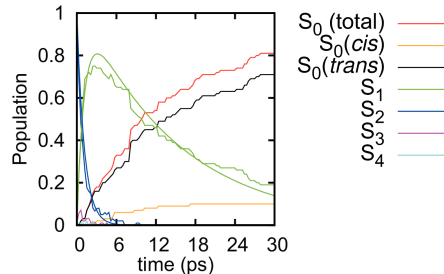
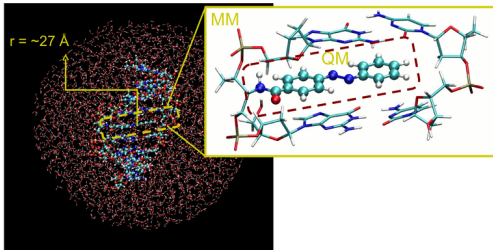
Padmabati Mondal; collaboration with M. Persico & co (Pisa)



- QM part: MOPAC / MM part: TINKER for RNA + ions + water
- electrostatic embedding / link atom scheme for covalent bonding

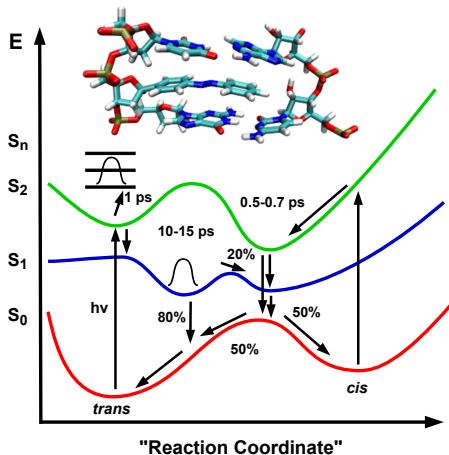
# Azobenzene & RNA: QM/MM + Surface Hopping

QM/MM = Quantum Mechanics/Molecular Mechanics



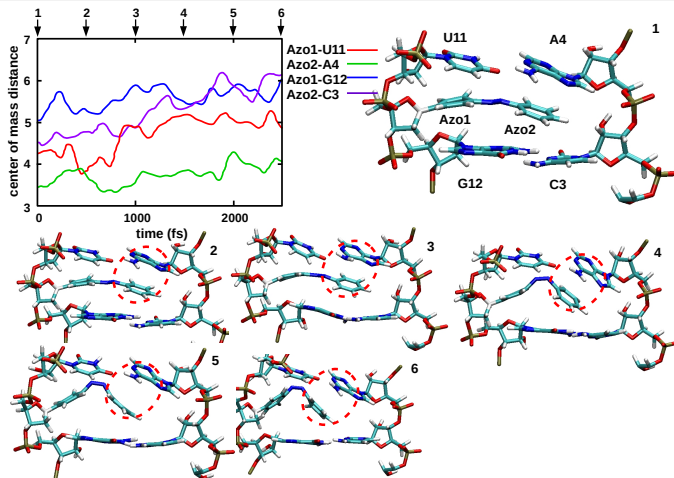
- *trans-cis* isomerization of azobenzene photoswitch triggers unfolding of RNA
- QM part: FOMO-SCF-CI (FOMO = Floating Occupation Molecular Orbital)
- MM part: TINKER (interfaced with MOPAC), Amber ff99sb force field
- QM/MM electrostatic embedding; connection atom scheme for covalent linkage
- initial excitation to  $S_2(\pi - \pi^*)$
- quantum yield  $\Phi_{\text{trans-cis}} = 10\%$ ,  $\Phi_{\text{cis-trans}} = 50\%$
- **comparatively slow *trans-cis* isomerization: ca. 20 ps instead of 300 fs (gas phase)**

## Modified dynamics in RNA environment



- rapid decay from "bright"  $S_2$  state to  $S_1$  state
- long  $S_1$  lifetime, presumably due to stacking interactions

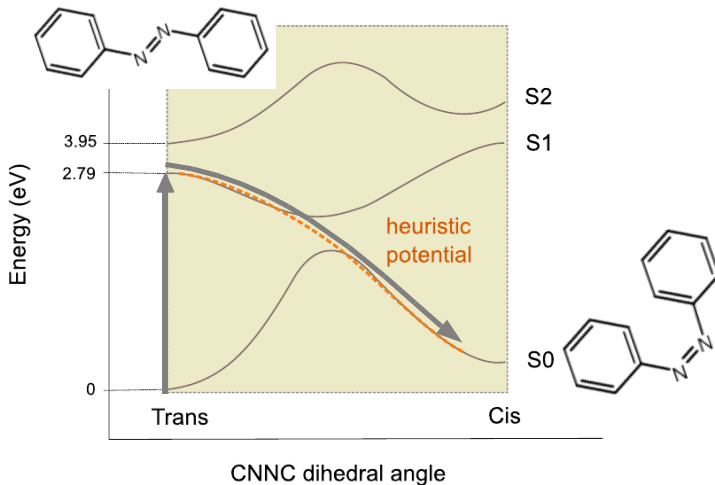
## How exactly does the dynamics happen?



- stacking interactions remain conserved as much as possible
- trajectory switches between  $S_1$  and  $S_0$  around  $t \sim 1.5$  ps

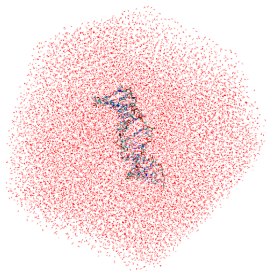
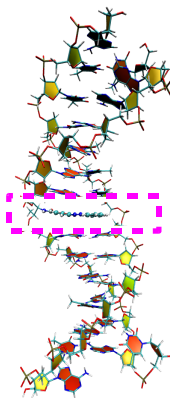
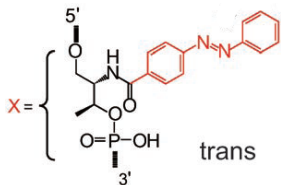
# Drastic simplification: isomerisation via effective potential

Biswas, Burghardt, Biophys. J., 107, 932 (2014)





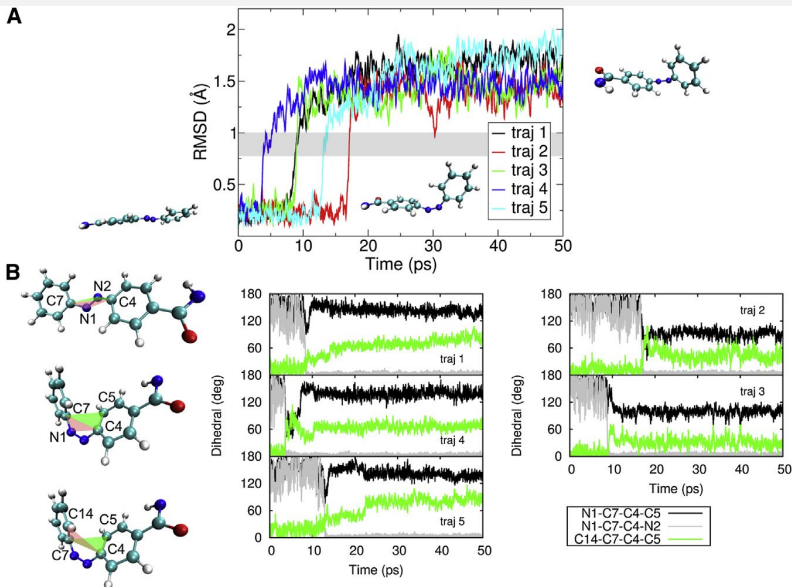
## Simulation Set-Up



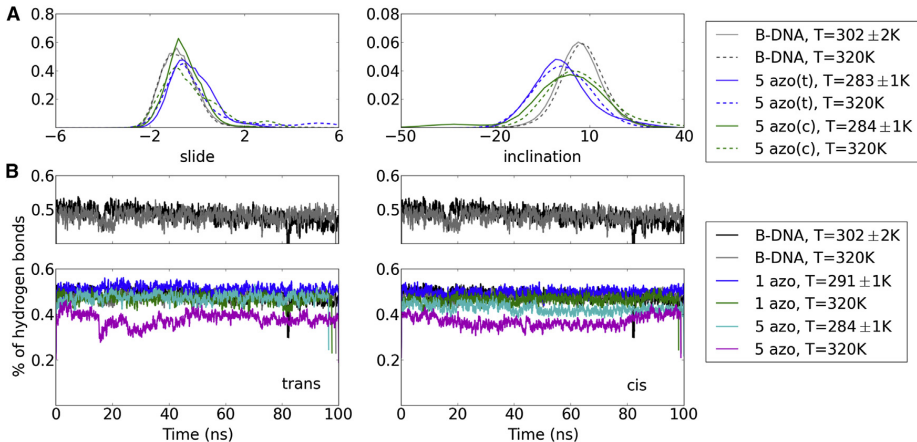
- 5'TAAGAAGXGAGATAT-3' sequence
- azobenzene + 14 mer DNA  
29357 atoms
- L-threoninol linkage
- AMBER99 force field  
with parambsc0 correction
- NAMD program
- 26 Na<sup>+</sup> ions
- 2 trans, 2 cis trajectories of  
duration 50 ns each

# Trajectory dynamics

Biswas, Burghardt, Biophys. J., 107, 932 (2014)



## DNA Destabilization

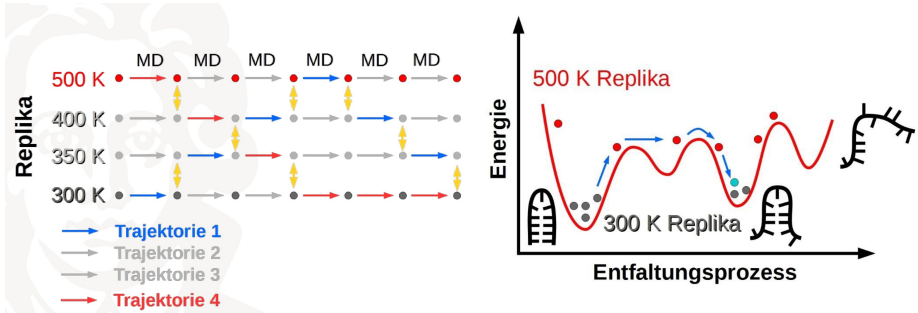


- distribution of helical parameters (slide, inclination) changes noticeably as a function of azobenzene attachment
- in this case, cis and trans forms do not show a significant difference

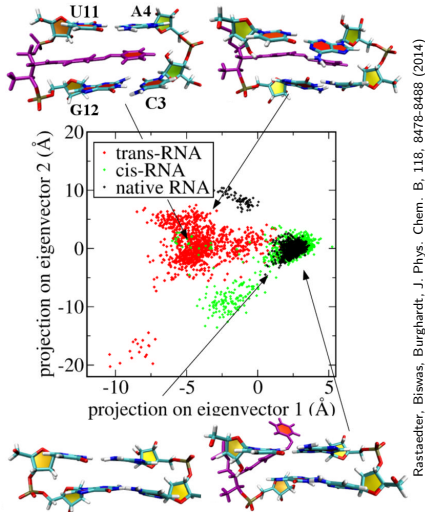
## Replica Exchange MD (REMD) simulations

- enhanced sampling technique: “watch” the unfolding of the double helix!
- REMD: run multiple isothermal MD simulations in parallel at a sequence of increasing temperatures and **intermittently swap** between temperatures

[Sugita, Okamoto, 1999]

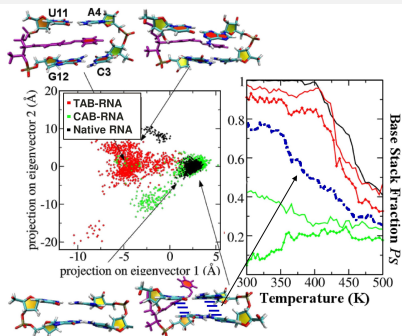
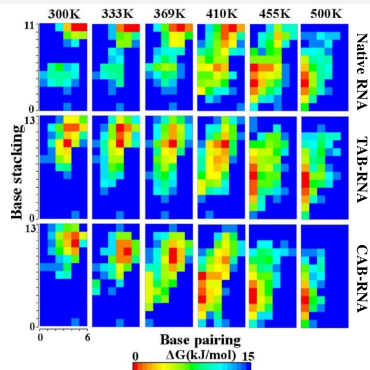


## Destabilization of Double Strand Structure



- several conformational ensembles for *trans* form (red)
- single conformational ensemble for *cis* form (black), close to native RNA → *cis* azobenzene has flipped sideways out of the double strand!
- degree of destabilization depends on several factors:
  - stacking interactions
  - hydrogen bonding
  - properties of the linker
  - presence of abasic site

# Free Energy Landscape and RNA Melting (RNA tetraloop hairpin, D-threoninol linker)



Rastaedt, Biswas, Burghardt, J. Phys. Chem. B, 118, 8478-8488 (2014)

- unfolding more rapid for the trans/cis substituted species than for native RNA
- folding parameter  $Q(t) = \frac{1}{2}(P_H(T) + P_S(T))$
- estimated melting temperatures  $T_m$ : cis/trans/native:  $382^\circ/418^\circ/427^\circ$
- scaled values  $T_m^{\text{scaled}}$ : cis/trans/native:  $305^\circ/334^\circ/342^\circ$

## Bottom line: multiscale methods needed!

